## ORGANIC LETTERS

2003 Vol. 5, No. 20 3627-3630

## Synthesis of Saponins Using Partially Protected Glycosyl Donors

Yuguo Du,\*,† Guofeng Gu,† Guohua Wei,† Yuxia Hua,† and Robert J. Linhardt\*,‡

Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing 100085, China, and Departments of Chemistry, Biology, and Chemical and Biological Engineering, Rensselaer Polytechnic Institute, Troy, New York 12180

duyuguo@mail.rcees.ac.cn; linhar@rpi.edu

Received July 21, 2003

## **ABSTRACT**

A new class of glycosyl donors having unprotected 2- and 2,4-hydroxyl groups were investigated under the standard glycosylation conditions. This approach was shown to be generally effective for the synthesis of alkyl and steroidal glycosides. A natural saponin, containing 2,4-branched oligosaccharide, was prepared in 35% overall yield in four straightforward sequential reactions by taking advantage of these partially protected donors.

Recent investigations in glycobiology have revealed important roles for many glycoconjugates in the immune response, viral and bacterial infection, cell regulation, differentiation, development, inflammation, cell adhesion, and many other inter- and intracellular communication processes. Saponins, a structurally and biologically diverse class of glycosides of steroids and triterpenes, are major components in traditional Chinese medicines and represent important examples of glycoconjugates with promising pharmaceutical and biological activities.<sup>2</sup> Dioscin (diosgenyl 2,4-di-*O*-α-L-rhamnopyranosyl- $\beta$ -D-glucopyranoside), polyphyllin D (diosgenyl  $\alpha$ -Lrhamno-pyranosyl- $(1\rightarrow 2)$ - $[\alpha$ -L-arabinofuranosyl- $(1\rightarrow 4)]$ - $\beta$ -D-glucopyranoside), and balanitin 7 (diosgenyl α-L-rhamnopyranosyl- $(1\rightarrow 2)$ - $[\beta$ -D-xylopyranosyl- $(1\rightarrow 3)$ - $\beta$ -D-glucopyranosyl- $(1\rightarrow 4)$ ]- $\beta$ -D-gluco-pyranoside) display cardiovascular, antifungal, and antitumor activities.<sup>3</sup> Moreover, the rhamnose moiety of solamargine (solasodinyl 2,4-di-O-αL-rhamnopyranosyl- $\beta$ -D-glucopyranoside) plays a crucial role in triggering cell death by apoptosis.<sup>4</sup> These bioactive saponins are comprised of a 2,4-branched oligosaccharide moiety, as are N-linked oligosaccharides and many plant polysaccharides.<sup>5</sup> Thus, because of their biological functions and also their unique 2,4-dibranched chain structures, the efficient synthesis of these steroidal glycosides deserves extensive exploration.

Substantial effort has been devoted to the development of novel glycosylation reactions as strategies to access natural glycoconjugate structures.<sup>6</sup> Saponins, having 2,4-branched oligosaccharides, have been traditionally assembled in three ways.<sup>7</sup> In the first approach, the reducing end sugar unit is coupled to the C-3 of a steroid or triterpene, next protection group manipulation is performed on the sugar residue, usually

<sup>†</sup> Chinese Academy of Sciences.

<sup>‡</sup> Rensselaer Polytechnic Institute.

<sup>(1)</sup> For reviews see: (a) Varki, A. Glycobiology **1993**, 3, 97. (b) Lee, Y. C.; Lee, R. T. Acc. Chem. Res. **1995**, 28, 321.

<sup>(2)</sup> Hostettmann, K.; Marston, A. Saponins; Cambridge University Press; New York, 1995.

<sup>(3) (</sup>a) Nakano, K.; Murakami, K.; Takaishi, Y.; Tomimatsu, T.; Nohara, T. Chem. Pharm. Bull. 1989, 37, 116. (b) Hufford, C. D.; Liu, S.; Clark, A. M. J. Nat. Prod. 1988, 51, 94. (c) Liu, C.; Chen, Y. Acta Pharm. Sinica 1984, 19, 799. (d) Namba, T.; Huang, X.; Shu, Y.; Huang, S.; Hattoti, M.; Kakiuchi, N.; Wang, Q.; Xu, G. Planta Med. 1989, 55, 501. (e) Zhou, J. Pure Appl. Chem. 1989, 61, 457.

<sup>(4)</sup> Chang, L.-C.; Tsai, T.-R.; Wang, J.-J.; Lin, C.-N.; Kuo, K.-W. *Biochem. Biophys. Res. Commun.* **1998**, 242, 21.

4,6-benzylidination followed by blocking the 3-hydroxyl group to give the glycosyl acceptor containing a free 2-hydroxyl group. Glycosylation at this 2-hydroxyl group, followed by selective opening of the 4,6-benzylidene, affords a free 4-hydroxyl group, which is further glycosylated to furnish a 2,4-branched saponin. The disadvantages of this method are that it involves a lengthy and low-efficiency synthesis. This is especially problematic when the aglycone is expensive or available only in limited quantities. In the second approach, a more highly convergent synthesis of saponin is carried out using a suitably modified monosaccharide donor with a participatory C-2 acyl protecting group to ensure the  $\beta$ -bond formation. However, the subsequent removal of C-2 acyl protecting groups from saponin derivatives, to expose the free 2-hydroxyl group for glycosylation, can be difficult.7d In the third approach, the 2,4-branched oligosaccharide is first prepared and then condensed with aglycone in the final step. Unfortunately, glycosylation with such an oligosaccharide results in decreased neighboring group participation and often generates  $\alpha,\beta$ -mixtures.<sup>7g</sup> With these difficulties in mind, we speculated that a partially protected glycosyl donor could be used to shorten the total synthesis of saponins.8

Model studies were first carried out on the preparation of alkyl glycosides using partially protected sugar donors and alkyl alcohol acceptors (Table 1) in  $CH_2Cl_2$  at -42 °C under

**Table 1.** Alkyl Glycoside Synthesis Using Partially Protected Glycosyl Donor

entry	donor	acceptor	product	yield
1	BzO OH BzO OH S-	$HOCH_2(CH_2)_5N_3$ $<$ <b>2</b>	BzO OH BzO OH	× 80%
2	BZO OH OBZ	< n-C <sub>8</sub> H <sub>17</sub> OH <sub>B</sub>	OH OBZ OH 6	<sub>CH₃</sub> <sup>a</sup> 83%
3	Ph O S-	∠ HOCH₂(CH₂)₅OTBS  8	Ph O OH	<sup>b</sup> 42% ∕∕отвѕ
a <b>A</b>	7	f a isomer was also is	9	220/ of the

 $^a$  A 7% yield of α isomer was also isolated.  $^b$  Including 32% of the  $\beta$ -isomer and 10% of the α-isomer.

promotion with N-iodosuccinimide (NIS) and trimethylsilyl trifluoromethanesulfate (TMSOTf). We were pleased to discover that mannopyranosyl thioglycoside  $\mathbf{1}$ , containing an unprotected hydroxyl group on C-2, still acted as an excellent glycosyl donor to afford  $\alpha$ -glycoside  $\mathbf{3}$  in high yield (80%).

No trace of self-condensed disaccharide was detected in our experiments. More impressively, when the galactopy-

(6) For reviews, see: (a) Toshima, K.; Tasuta, K. Chem. Rev. 1993, 93, 1503. (b) Schmidt, R. R.; Kinzy, W. Adv. Carbohydr. Chem. Biochem. 1994, 50, 21. (c) Garegg, P. J. Adv. Carbohydr. Chem. Biochem. 1997, 52, 179.

**Table 2.** Saponin Synthesis Using Partially Protected Glycosyl Donors

Donors						
entry	donor	acceptor	product	yield		
1	HO OBZ BZO O S	HO 10	HO OBZ BZO Odios	54%		
2	TBSO 12 OH	10	TBSO Odios	<sup>a</sup> 59%		
3	Ph O S	10	complex	n.d.		
4	FmocO S S	10	FmocO Odios	75%		
5	HO OBZ BZO OH	10	HO Odios BzO Odios	<sup>b</sup> 54%		
6	BnO OBn BnO SEt	10	BnO Odios 19 OH	80%		
7	Bzo OH	но 21	BzO Ochol	68%		
8	O OTBS O S-(	21	OTBS OCCHOI	<sup>c</sup> 55%		
9	25 OH	21	26 <sup>OH</sup>	<sup>d</sup> 45%		

 $^a$  An additional 6% of the  $\alpha$ -isomer was isolated.  $^b$   $\alpha$ :  $\beta$  = 1:2.  $^c$   $\alpha$ :  $\beta$  = 2:3.  $^d$  An additional 14% yield of the disaccharide saponin derivative was also isolated.

ranosyl thioglycoside donor **4**, containing 2,4-dihydroxyl groups, was subjected to similar reaction conditions, octyl  $\beta$ -D-galactopyranoside **6** was obtained in a yield of 83% ( $\alpha$ :  $\beta = 10:1$ ). Furthermore, 2,3-dihydroxyl donor **7** afforded a modest (42%) yield of galactopyranoside **9** as a 1:3  $\alpha/\beta$  mixture.

Encouraged by these preliminary results, we next turned our attention to saponin synthesis (Table 2). Condensation of donor **4** with diosgenin **10** in CH<sub>2</sub>Cl<sub>2</sub> at -42 °C under NIS-TMSOTf promotion afforded a 54% isolated yield of  $\beta$ -glycoside **11**. A doublet at 4.53 ppm (J = 7.7 Hz) in <sup>1</sup>H NMR spectrum clearly demonstrated the pure  $\beta$ -configuration

Org. Lett., Vol. 5, No. 20, 2003

<sup>(5) (</sup>a) Iorizzi, M.; De Marino, S.; Zollo, F. Curr. Org. Chem. 2001, 5, 951. (b) Bedir, E.; Khan, I. A. J. Nat. Prod. 2000, 63, 1699. (c) Yin, J.; Kouda, K.; Tezuka, Y.; Le Tran, Q.; Miyahara, T.; Chen, Y.; Kadota, S. J. Nat. Prod. 2003, 66, 646. (d) Nakamura, T.; Komori, C.; Lee, Y.-Y.; Hashimoto, F.; Yahara, S.; Nohara, T.; Ejima, A. Biol. Pharm. Bull. 1996, 19, 564. (e) Miyamura, M.; Nakano, K.; Nohara, T.; Tomimatsu, T.; Kawasaki, T. Chem. Pharm. Bull. 1982, 30, 712. (f) Akhov, L. S.; Musienko, M. M.; Piacente, S.; Pizza, C.; Oleszek, W. J. Agric. Food Chem. 1999, 47, 3193. (g) Dwek, R. A. Chem. Rev. 1996, 96, 683.

<sup>(7) (</sup>a) Lahmann, M.; Gybäck, H.; Garegg, P. J.; Oscarson, S.; Suhr, R.; Thiem, J. Carbohydr. Res. 2002, 337, 2153. (b) Yu, H.; Yu, B.; Wu, X.; Hui, Y.; Han, X. J. Chem. Soc., Perkin Trans. J 2000, 1445. (c) Cheng, M.; Wang, Q.; Tian, Q.; Song, H.; Liu, Y.; Li, Q.; Xu, X.; Miao, H.; Yao, X.; Yang, Z. J. Org. Chem. 2003, 68, 3658. (d) Deng, S.; Yu, B.; Hui, Y., Yu, B.; Han, X. Carbohydr. Res. 1999, 317, 53. (e) Deng, S.; Yu, B.; Hui, Y. Tetrahedron Lett. 1998, 39, 6511. (f) Li, B.; Yu, B.; Hui, Y.; Li, M.; Han, X.; Fung, K.-P. Carbohydr. Res. 2001, 331, 1. (g) Ikeda, T.; Miyashita, H.; Kajimoto, T.; Nohara, T. Tetrahedron Lett. 2001, 42, 2353.

<sup>(8)</sup> Plante, O.; Palmacci, E. R.; Andrade, R. B.; Seeberger, P. H. J. Am. Chem. Soc. 2001, 123, 9545.

<sup>(9)</sup> **General Procedure.** To a mixture of thioglycosyl donor (1 mmol) and ROH (1 mmol) in anhydrous dichloromethane (2 mL) at  $-42\,^{\circ}\mathrm{C}$  were added 1.1 mmol of NIS and catalytic amount of TMSOTf (0.1 equiv) with  $N_2$  protection. The reaction mixture was stirred under these conditions for 45 min, at which time TLC indicated the completion of the reaction. The mixture was then neutralized with  $Et_3N$  and concentrated to dryness. The residue was subjected to column chromatography on silica gel with petroleum ether/EtOAc (6/1-3/1) as the eluent to give the desired product.

of 11. The  $\alpha$ -isomer might also be generated in this reaction, as the presence of inseparable contaminants did not permit us to rule out its formation. The 3,6-disilylated donor 12 gave an easily separable  $\beta$ -glycoside 13 (59%), together with a 6% yield of the  $\alpha$ -product under the same reaction conditions. The 4,6-benzylidenated donor 7 produced a complex product mixture. In contrast, the corresponding C-3 protected donors 14 and 20 provided 15 and 22, respectively, in better yields and excellent regioselectivities, indicating that appropriate protection of the 3-hydroxyl group is critical for the effective application of this type of glycosyl donor. In parallel experiments, the stereochemical outcomes of 3,4isopropylidenated donors were greatly influenced by the substitution on C-6. For example, 6-silvlated donor 23 gave cholesterol saponin 24 as an  $\alpha,\beta$ -mixture with low stereoselectivity, but 6-deoxy donor 25 afforded 45% yield of  $\beta$ -glycoside **26**. When 2,4-unprotected glucosyl donor **16** was coupled with diosgenin 10 under the same reaction conditions, only a 36% yield of  $\beta$ -glycoside 17 was obtained, much lower than that obtained with the corresponding galactosyl donor 4. Interestingly, using a partially benzylated glucosyl donor 18 significantly improved the yield (80%) and gave  $\beta$ -glycoside **19** as the sole product.

Coupling reactions between sugar residues were also investigated using thioglycoside donors with unprotected 2-hydroxyl or 2,4-hydroxyl groups (Table 3). When glucosyl

**Table 3.** Oligosaccharide Synthesis Using Partial Protected Glycosyl Donor

entry	donor	acceptor	product	yield
1	18	27 OH	BnO HO O	77%
2	4	27	HO OB Z BZO HO O O 29	<sup>a</sup> 50%
3	23	MeO OH	OTB S Ho Meo 31 Meo	<sup>b</sup> 57%
4	18	BnO OMe	BnO OBn OBn OMe	20%
5	18	BZO OME	complex	n.d.

 $^a$  An α:  $\beta$  = 1:2 mixture with additional trisaccharide (15%) was isolated.  $^b$  An α:  $\beta$  = 2:3 mixture.

donor 18 was reacted with 1,2:3,4-di-O-isopropylidene- $\alpha$ -D-galactopyranose (27) in CH<sub>2</sub>Cl<sub>2</sub> at -42 °C in the presence of NIS-TMSOTf, a 77% yield of disaccharide 28 was isolated. However, a similar reaction between 4 and 27 gave only 50% yield of the  $\alpha$ , $\beta$ -mixture 29. An inseparable  $\alpha$ , $\beta$ -mixture of glycoside 31 ( $\beta$ :  $\alpha$  = 3:2) was also obtained in 57% yield on glycosylation of 30 with 23. When sugar acceptors containing secondary hydroxyl groups (32 and 34) were examined, low yields or complex product mixtures were obtained (Table 3, entries 4 and 5).

The formation of the predominantly  $\beta$ -product can be rationalized as shown in Scheme 1.<sup>10</sup> When glycosylation

with thioglycoside **A** was promoted using NIS and TMSOTf, two reactive intermediates **B** and **C** could be generated. The 1,2-anhydrosugar intermediate **C** is formed through the intermolecular ring closure of **B**. Stereoselective opening of intermediate **C** would afford  $\beta$ -product **E**, while the  $\alpha$ , $\beta$ -mixture **D** would be obtained through  $\alpha$ - and  $\beta$ -attack on oxocarbenium **B**.

To ascertain the efficiency of our new synthetic method, we next applied it to the synthesis of diosgenyl  $\alpha$ -L-rhamnopyranosyl- $(1\rightarrow 2)$ - $[\beta$ -D-glucopyranosyl- $(1\rightarrow 4)]$ - $\beta$ -D-galactopyranoside, <sup>5f</sup> a potent drug candidate used to decrease the cholesterol level in serum.

Compound **4**, containing unprotected 2,4-hydroxyl groups, was prepared from commercially available IPTG **35** according to the method described by Chan (Scheme 2).<sup>11</sup> This

 $^a$  Reaction conditions: (a) BzCl, Pyr, −10 °C, 70%. (b) **10**, NIS, TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>, −42 °C, 54%. (c) **36**, TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>, −42 °C; then **37**, TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 68.7% for one-pot reaction. (d) Aqueous 1 N NaOH, MeOH, 95%.

donor was condensed with diosgenin 10 in  $CH_2Cl_2$  at -42 °C in the presence of NIS-TMSOTf, to afford the desired

Org. Lett., Vol. 5, No. 20, 2003

<sup>(10) (</sup>a) Leeuwenburgh, M. A.; Timmers, C. M.; van der Marel, G. A.; van Boom, J. H.; Mallet, J.-M.; Sinay, P. G. *Tetrahedron Lett.* **1997**, *38*, 6251. (b) Du, Y.; Kong, F. *J. Carbohydr. Chem.* **1995**, *14*, 341.

 $\beta$ -glycoside 11 in 54% isolated yield. Compound 11 was glycosylated in one-pot with rhamnopyranosyl trichloroace-timidate 36 at -42 °C in the presence of TMSOTf, followed by glucopyranosyl imidate 37 at 0 °C, to afford the protected trisaccharide saponin derivative 38 (68.7% from 11). Natural saponin 39 was then readily obtained by deacylation with aqueous 1 N NaOH in MeOH (95%). Remarkably, this complex natural saponin was prepared in four simple steps and in 35% overall yield.

In conclusion, an efficient and practical method has been developed for the preparation of saponins having 2,4-branched oligosaccharide moieties. The key to this chemistry is the use of partially unprotected thioglycosides as glycosyl donors. This results in significantly simplified protecting group manipulation and oligosaccharide assembly. The approach described is general and effective for alkyl and

(11) Jiang, L.; Chan, T.-H. J. Org. Chem. 1998, 63, 6035.

steroidal glycoside synthesis. More importantly, the application of this method with combinatorial chemistry might be useful as an efficient entry into libraries of more complex glycoconjugates.<sup>12</sup>

**Acknowledgment.** This work was supported by NNSF of China (29972053), RCEES of CAS, and NIH of the U.S. (HL62244).

Supporting Information Available: Preparation and physical data for compounds 3, 6, 9, 11, 13, 15, 17, 19, 22, 26, 28, 29, 33, 38, and 39. This material is available free of charge via the Internet at http://pubs.acs.org.

## OL035353S

3630 Org. Lett., Vol. 5, No. 20, 2003

<sup>(12)</sup> Marcaurelle, L. A.; Seeberger, P. H. Curr. Opin. Chem. Biol. 2002, 6, 289.